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Research Article

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In-vitro Antibacterial Efficacy of Solvent Extracts of Leaves of Bauhinia racemosa Lam. (Caesalpiniaceae) Against Enteric Bacterial Pathogens

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ABSTRACT

Phytochemical screening of the plant leaves reveals the presence of carbohydrates, alkaloids, flavonoids, steroids, and tannins. Petroleum ether extract, chloroform extract, ethyl acetate extract and methanol extracts of leaves of *Bauhinia racemosa* Linn. were prepared and antibacterial activity were studied by disc diffusion method against certain enteric bacterial pathogens such as *Escherichia coli, Staphylococcus aureus, Klebsiella pneumonia, Enterobacter aerogenes, Pseudomonas aeruginosa, Salmonella typhimurium, Salmonella typhi, Staphylococcus epidermidis* and *Proteus vulgaris*. The Methanol extracts had wide range of antibacterial activity against enteric bacterial pathogens than the petroleum ether extract, where as ethyl acetate extract were slightly higher antibacterial activity than chloroform extract. Antibacterial activity of various extract of leaves of *Bauhinia racemosa* was carried in attempt to develop a new pharmaceutical drug from natural origin for prevention of enteric infection.

Keywords: Antibacterial activity, *Bauhinia racemosa*, enteric bacterial pathogens.

INTRODUCTION

The use of plant compounds for pharmaceutical purposes has gradually increased in India. About 80% of individuals from developed countries use traditional medicine, which involves compounds derived from medicinal plants. [1] Therefore, such plants should be investigated to better understand their properties, safety and efficiency. The use of plant extracts and phytochemicals, both with known antimicrobial properties, can be of great significance in therapeutic treatments. [2] Hence, studies involving the use of plants as therapeutic agents should be emphasized, especially those related to the control of antibiotic resistant microbes. [3] The plant Bauhinia racemosa (L). belongs to the Caesalpiniaceae family. It occurs frequently in India, Ceylon, China, and Timor. The stem bark of the plant is an astringent and is used in the treatment of headache, fever, skin diseases, tumors, blood diseases, dysentery, and diarrhea. [4] The fresh flower buds of the plant showed antiulcer activity. [5-6]

Enteric or diarrhoeal infections are major public health problems in developing countries and contribute to the death

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of 3.3 to 6.0 million children annually. Enteric bacteria comprised of *Salmonella* sp., *Shigella* sp., *Proteus* sp., *Klebsiella* sp., *E. coli, Pseudomonas* sp., *Vibrio cholerae* and *Staphylococcus aureus* which are major etiologic agents of sporadic and epidemic diarrhea both in children and adults. ^[7] Therefore the review revealed that the leaves of *Bauhinia racemosa* were used in various metabolic disorders, but far their antibacterial properties were not demonstrated. Hence attempt was made to find out the antibacterial properties of leaves of *Bauhinia racemosa* against enteric bacterial pathogens.

MATERIALS AND METHODS

Plant materials

Fresh plant of *Bauhinia racemosa* (L). were collected from local region of Ahmednagar District in India. The leaves were identified by Mr. P. S. N. Rao, Join Director, Botanical survey of India (BSI), Koregaon road, Pune.

Preparation of extracts

1.5 kg of the plant material in each batch was exhaustively extracted by soxhlet extraction method using Petroleum ether, Chloroform, Ethyl acetate and Methanol. The solvent used in each batch was recovered under pressure until dry extracts were obtained and then labeled and stored separately at 4°C in amber colored airtight bottles.

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Phytochemical Screening of Plant materials

The presence of saponins, tannins, carbohydrates, alkaloids, flavonoids glycosides, steroids, proteins and alkaloids, were detected by simple qualitative methods. ^[9]

Bacterial cultures

The standard pathogenic bacterial cultures were procured from IMTECH, Chandigarh, India and used in the present study (Table 1). The bacterial cultures were rejuvenated in Mueller- Hinton broth (Hi-media laboratories, Mumbai, India) at 37°C for 18h and then stocked at 4°C in Mueller-Hinton Agar. The inoculum size of the bacterial culture was standardized according to the National committee for Clinical Laboratory Standards (NCCLS, 2002) guideline. The pathogenic bacterial culture was inoculated into sterile Nutrient broth and incubated at 37°C for 3h until the culture attained a turbidity of 0.5 McFarland units. The final inoculum size was standardized to 10^5 CFU/mL with the help of SPC and Nephlo-turbidometer.

Table 1: Bacterial cultures used in study (IMTECH, Chandigarh, India)

Bacterial Pathogens	MTCC Number						
Proteus vulgaris	426						
Staphylococcus epidermidis	435						
Staphylococcus aureus	96						
Escherichia coli	739						
Pseudomonas aeruginosa	424						
Klebsiella pneumoniae	109						
Salmonella typhi	733						
Enterobacter aerogenes	111						
Salmonella typhimurium	98						

Preparation of disc for antibacterial activities

The Petroleum ether, Chloroform, Ethyl acetate and Methanol extracts were prepared in their respective solvents and the sterile blotting paper disc (10 mm) were soaked in the diluted extract in such concentration that the amount of solution absorbed by each disc was 1mg, 2mg, 3mg, 4mg, 5mg of each extracts of *Bauhinia racemosa*. The prepared disc were dried in controlled temperature to remove excess of solvent and used in study.

Antibacterial activity using disc diffusion method

The modified paper disc diffusion [10] was employed to determine the antibacterial activity of The Petroleum ether, Chloroform, Ethyl acetate and Methanol extracts of leaves of *Bauhinia racemosa*. Turbidity of inoculums was matched with McFarland turbidity standard. Inoculums were spread over the Nutrient agar plate using a sterile cotton swab in order to get a uniform microbial growth. Then the prepared antibacterial disc were placed over the lawn and pressed slightly along with positive and negative controls. Ampicillin 10 mcg/disc (Hi-Media, Mumbai) were used as positive control while disc soaked in sterile distilled water or various organic solvents and dried were placed on lawns as negative control. The plates were incubated for 18h at 37°C. The antibacterial activity was evaluated and diameters of Table 2: Phytochemical analysis of *Bauhinia racemosa* leaves.

inhibition zones were measured. Experiment was carried out in triplicate and the averages diameter of zone of inhibition was recorded. The antibacterial activity was classified as strong (>20mm), moderate (16-19mm) and mild (12-15mm) and less than 12mm was taken as inactive.

RESULTS AND DISCUSSION

The photochemical investigation (Table 2) of the various solvent extract of leaves of *Bauhinia racemosa* showed the Petroleum ether extract to only alkaloids and steroids which, occurring in higher concentration. The chloroform extract and ethyl acetate extract content similar photochemical constitutes, alkaloids and steroids in higher concentration, Carbohydrates, Flavonoids, glycosides, saponins. Methanol extract contented Alkaloids, Steroids, Flavonoids, Saponins and Tannins, but did not contended any proteins.

According to antibacterial profile (Table 3), maximum inhibitory effect of the Petroleum ether extract observed only on Staphylococcus epidermidis, Staphylococcus aureus, Salmonella typhi and moderate antibacterial against Escherichia coli, Pseudomonas aeruginosa, Enterobacter aerogenes, Salmonella typhimurium but mild inhibitory effect on Proteus vulgaris. Chloroform extract showed strong antibacterial effect against Staphylococcus epidermidis and Staphylococcus aureus and moderate antibacterial against Proteus vulgaris, Escherichia coli, Enterobacter aerogenes, Salmonella typhi and Salmonella typhimurium but mild effect on Pseudomonas aeruginosa. Ethyl acetate extract showed maximum inhibitory effect on Staphylococcus epidermidis, Staphylococcus aureus Escherichia coli, Pseudomonas aeruginosa, Enterobacter aerogenes and Salmonella typhimurium, but moderate effect on Proteus vulgaris and Salmonella typhi. Methanol extract showed maximum inhibitory effect on Staphylococcus aureus, Proteus vulgaris, Staphylococcus epidermidis, Pseudomonas aeruginosa, Salmonella typhi, Salmonella typhimurium, but moderate inhibitory effect on Escherichia coli, Enterobacter aerogenes.

The result of the antibacterial assay show promising evidence for the antibacterial effect of leaves of *Bauhinia racemosa*. Form the above evidence, it is clear that plant extracts have great potential as antibacterial compounds against enteric pathogens and that they can be used in the treatment of enteric infectious. It is hoped that this study would lead to the establishment of some compounds that could be used to formulate new and more potent antimicrobial drugs of natural origin.

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S. No	Phytochemical Constitutes	Petroleum ether extract	Chloroform extract	Ethyl acetate extract	Methanol extract
1	Alkaloid	+	+	+	+
2	Flavonoids	-	-	+	+
3	Carbohydrates	+	+	-	-
4	Glycosides	-	-	-	-
5	Saponins	-	-	-	+
6	Proteins	+	-	-	-
7	Steroids	-	-	+	-
8	Tannins	+	-	-	-

Where: + = the presence of constitute, - = the absence of constitutes

Table 3: Antibacterial activity of Bauhinia racemosa leaves extracts against enteric bacterial pathogens (Zone of inhibition of growth in mm, average of 3 readings)

readings)	The Petroleum ether extract				Chloroform extract				Ethyl acetate extract					Methanol extract						con					
Bacterial Pathogens	5mg /disc	4mg /disc	3mg /disc	2mg /disc	1mg /disc	5mg /disc	4mg /disc	3mg /disc	2mg /disc	1mg/disc	5mg /disc	4mg /disc	3mg /disc	2mg /disc	1mg/disc	5mg /disc	4mg/disc	3mg /disc	2mg /disc	1mg/disc	Pet ether	Chloroform	Ethyl acetate	Methanol	Ampicillir (10mcg)
P. vulgaris	15	13	12	-	-	17	15	12	-	-	19	17	15	15	12	20	17	15	13	12	-	-	-	-	16
S. epidermidis	23	20	17	15	13	21	19	17	16	14	24	22	20	17	15	26	18	20	18	14	-	-	-	-	25
Ŝ. aureus	22	21	18	17	16	24	22	20	18	16	20	19	17	15	14	22	19	18	16	13	-	-	-	-	24
E.coli	19	17	15	12	12	17	15	14	12	-	22	19	17	14	12	17	15	14	12	-	-	-	-	-	11
P. aeruginosa	16	15	14	13	12	15	14	13	-	-	21	17	15	14	13	20	17	16	15	13	-	-	-	-	16
S. typhi	21	20	17	14	12	18	16	15	14	13	18	17	16	15	14	22	20	18	15	14	-	-	-	-	18
E. aerogenes	17	15	15	13	12	18	16	15	13	11	22	19	17	14	13	17	15	13	13	12	-	-	-	-	14
S typhimurium	18	17	15	13	-11	18	17	14	12	_	2.0	17	17	15	14	22	20	17	14	12	_	_	_	_	19

REFERENCES

- Chopra RN, Nayar SL, Chopra IC. Glossary of Indian Medicinal Plants (Including the Supplement). Council of Scientific and Industrial Research, New Delhi, 1986.
- Kiritikar KR and Basu BD. Indian medicinal plants. 2nd edn. International Book Distribution Vol. III. Dehradun, 1987, pp.1664– 1666
- Parekh J, Nair R, Chanda S. Preliminary screening of some folkloric plants from Western India for potential antimicrobial activity. Indian J Pharmacol. 2005; 37: 408-409.
- Prakash A and Khosa RL. Chemical studies on *Bauhinia racemosa*. Current Science, 1976; 45: 705-707.
- El-Khatiba AS and Khaleel AE. Evaluation of some pharmacological properties of different extracts of *Bauhinia* racemosa leaf and *Bassia muricata* whole plant. Bulletin of the Faculty of Pharmacy, Cairo University, 1995; 33: 59-65.

- El-Hossary GA, Selim MA, Sayed AE and Khaleel AE. Study of the flavonoid content of *Bassia muricata* and *Bauhinia racemosa*. Bulletin of the Faculty of Pharmacy, Cairo University, 2000; 38: 93-97.
- World Health Organization, 5 th Programme Report, Programme for control of diarrhoeal diseases, Geneva. WHO Bulletin, 1985; 63: 557-772.
- Ballal M. Screening of medicinal plants used in rural folk medicine for treatment of diarrhea, 2005. Internet: http:// www.Pharmoinfo.net.
- Khandelwal KR. Preliminary photochemical screening, in: Practical Pharmacognosy Techniques and Experiments, Edn. 8, Nirali Publication, Pune, 20011, pp. 49-156.
- NCCLS (National Committee for Clinical Laboratory Standards), Performance Standards for antimicrobial susceptibility testing. 8th Informational Supplement. M100 S12. National Committee for Clinical Laboratory Standards, Villanova, Pa 2002.