

Contents lists available at UGC-CARE

International Journal of Pharmaceutical Sciences and Drug Research

[ISSN: 0975-248X; CODEN (USA): IJPSPP]

Available online at www.ijpsdronline.com



Research Article

Stability-indicating Reversed Phase-Ultra Performance Liquid Chromatography Method Development and Validation for Simultaneous Determination of Encorafenib and Binimetinib in Formulation

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ARTICLE INFO

Article history:

Received: 03 May, 2020 Revised: 12 August, 2020 Accepted: 25 August, 2020 Published: 30 September, 2020

Keywords:

Binimetinib, Encorafenib, Stability studies, Ultra performance liquid chromatography (UPLC), Validation.

DOI:

10.25004/IJPSDR.2020.120509

ABSTRACT

Simple, accurate, and the precise stability-indicating method was developed for the simultaneous estimation of the encorafenib (ECRB) and binimetinib (BMTB) in a dosage form by ultra performance liquid chromatography (UPLC). Chromatographic elution was processed through an HSS C18 (100 × 2.1 mm, 1.8 m) reverse phase column, and the mobile phase composition of 0.01N $\rm KH_2PO_4$ buffer (pH 3.5) and acetonitrile in the proportion of 55:45 was processed through a column at a flow rate of 1 mL/min. The temperature of the column oven was kept at 30°C, and the wavelength maximum of detection system was set to 294 nm. Retention times of ECRB and BMTB were found to be 0.767 and 1.13 minutes, respectively. Repeatability of the method was determined in the form of % relative standard deviation (RSD), and findings were 0.3 and 0.6 for ECRB and BMTB, respectively. The percentage recovery of the method was found to be 99.59 and 99.7% for ECRB and BMTB, respectively. The limit of detection (LOD) and the limit of quantification (LOQ) values obtained from regression equations of ECRB and BMTB were 0.51, 1.55 mg/mL and 1.47, 4.44 mg/mL, respectively. The regression equation of ECRB was y = 6,684x + 18,102, and BMTB was y = 13,118x + 2,159. Two analytes were subjected to acid, peroxide, photolytic, alkali, neutral, and thermal degradation studies, and the results shown that the percentage of degradation was found between 0.76 and 6.88%. Retention times and the total run time of two drugs were decreased, and the developed method was simple and economical. So, the developed method can be adopted in industries as a regular quality control test for the quantification of ECRB and BMTB.

INTRODUCTION

The ECRB is a drug component available in the market as Braftovi, utilized in the treatment of different melanomas. It belongs to a BRAF inhibitor that affects the key enzymes involved in the mitogen-activated protein kinase (MAPK) signaling path. This path of signaling takes place in many kinds of cancers, together with colorectal cancer and melanomas. [1,2] ECRB inhibits the ATP-competitive RAF kinase, downregulates cyclin-D1, and decreases ERK phosphorylation. [3-5] This stops the cell cycling process in phase-G1, prompting senescence without apoptosis. ECRB chemically designated as methyl[(2S)-1-{[4-(3-{5-chloro-2-fluoro-3-[(methylsulfonyl) amino] phenyl}-1-isopropyl-

1H-pyrazol-4-yl)-2-pyrimidinyl] amino}-2-propanyl] carbamate^[1,3] with molecular weight and formula of 540.011 g/mole and $C_{22}H_{27}ClFN_7O_4S$, respectively (Fig. 1).

BMTB (trade name Mektovi) selectively inhibits MEK, a central kinase in the tumor stimulating MAPK-path. Incongruous stimulation of the path has been shown to ensue in several cancers. [6] BMTB is a mitogen-activated protein kinase (MEK) inhibitor available orally, or, more specifically, an inhibitor of MAP2K. [7] MEK is part of the rat sarcoma (RAS) pathway, which is involved in cell proliferation and survival. MEK is upregulated in many forms of cancer. [8] BMTB, uncompetitive with ATP, binds to and inhibits the activity of MEK1/2 kinase, which has been

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Relevant conflicts of interest/financial disclosures: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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shown to regulate several key cellular activities, including proliferation, survival, and angiogenesis. BMTB chemically designated as 5-((4-bromo-2-fluorophenyl) amino)-4-fluoro-N-(2-hydroxyethoxy)-1-methyl-1H-benzo[d] imidazole-6-carboxamide $^{[6-8]}$ with molecular weight and formula of 441.23 g/mole and $C_{17}H_{15}{\rm Br}F_2N_4O_3$, respectively (Fig. 1).

ECRB and BMTB target two dissimilar kinases in the path of RAS-RAF-MEK-ERK. A combination of ECRB and BMTB results in superior anti-proliferating action *in vitro* in BRAF mutation-positive cell lines, compared with activity of any single drug alone. $^{[7]}$ In addition to the above, these two drug combinations acted to defer the emergence of resistance in BRAF-V600E mutant human melanoma xenografts in mice compared with the administration of any one drug alone. $^{[4,8]}$

The literature review unveils that very less UPLC-MS/MS^[9] and reverse-phase high performance liquid chromatographic (RP-HPLC)^[10] techniques have been established for the determination of ECRB and BMTB. Based on the reported HPLC methods, there is a need to develop a stability-indicating RP-UPLC method for the simultaneous estimation of ECRB and BMTB in bulk and dosage form.

MATERIALS AND METHODS

Chemicals and Reagents

The active pharmaceutical ingredient (API) of ECRB and BMTB were obtained from Spectrum Pharma Research Solutions, Hyderabad. HPLC-grade methanol and acetonitrile were procured from Merck Chemical Division, Mumbai, India. Potassium dihydrogen orthophosphate, orthophosphoric acid, sodium dihydrogen orthophosphate, and HPLC-grade water were bought from Rankem, Avantor Performance Material India Limited. Braftovi capsules and Mektovi tablets were obtained from a local pharmacy.

Chromatographic System

Liquid chromatographic UPLC system of Waters equipped with photodiode array detector (PDA), auto-sampling unit, and HSS C18 (100 \times 2.1 mm, 1.8 m) reverse phase column. The mobile phase combination of 0.01N KH $_2$ PO $_4$ buffer (pH 3.5) and acetonitrile in the ratio of 55:45 was pumped through a column at a flow rate of 1 mL/min. The column oven temperature was maintained at 30°C, and the detection wavelength was processed at 294 nm. The

Fig. 1: Chemical structures of a) ECRB; b) BMTB

integration of output signals was monitored and processed by waters Empower software-2.0.

Diluents

Depending upon the solubility of the drugs, diluent was optimized. Initially dissolved in methanol and diluted with acetonitrile and water (50:50).

Preparation of Standard Stock Solutions

Exactly weighed 90 mg of ECRB and 9 mg of BMTB poured into two 50 mL volumetric flasks alone. 10 mL of diluent was added and vortexed for 10 minutes. Flasks were made up of diluent and marked as standard stock solutions 1 and 2 (1,800 µg/mL of ECRB and 180 µg/mL BMTB). From each stock solution, 1 mL was pipetted out and poured into a 10 mL volumetric flask, and the final volume was made up to mark with diluent to get 180 µg/mL of ECRB and 18 µg/mL of BMTB.

Preparation of Sample Stock Solutions

5 tablets were weighed, and the average weight of each tablet was calculated. The weight equivalent to one tablet was transferred into a 500 mL volumetric flask, and 25 mL of diluent was added and sonicated for 25 minutes. Further, the volume was made up with diluent and filtered through a 0.45 μ filter (900 μ g/mL of ECRB and 90 μ g/mL of BMTB). 2 mL of the resultant solution was poured into a 10 mL volumetric flask and made up with diluent (180 μ g/mL of ECRB and 18 μ g/mL of BMTB).

Preparation of Buffer

Accurately weighed 1.36 gm of potassium dihydrogen orthophosphate in a 1,000 mL of volumetric flask and added about 800 mL of milli-Q water. The resulting solution was subjected to sonication for 10 minutes, make up the volume with water, and then adjust the pH to 3.5 with 0.1% orthophosphoric acid solution.

Method Validation

The developed method for ECRB and BMTB was subjected to validation for the parameters, like system suitability, robustness, linearity, LOD, precision, LOQ, and accuracy as per the ICH guidelines. [11-15]

RESULTS

Method Development and Optimization

We tried different mobile phase combinations with methanol, water, acetonitrile, and buffer. At all the combinations, the resulting chromatograms got a poor resolution, theoretical plates, and peak shape. [15-17] Finally, excellent chromatographic efficiency parameters were obtained with the mobile phase composition of 0.01N KH₂PO₄ buffer (pH 3.5), and acetonitrile in the ratio of 55:45 %v/v pumped through an HSS C18 (100 × 2.1 mm, 1.8 m) reverse phase column, at a flow rate of 1 mL/min. The

column oven temperature was maintained at 30° C, and the detection wavelength was processed at 294 nm. Based on the solubility, all the dilutions were made with acetonitrile and water in the ratio of 50:50 %v/v. Retention times of ECRB and BMTB were found to be 0.767 and 1.13 minutes, respectively. An injection volume of 0.3 mL was infused through a UPLC system to get better performance.

System Suitability

The system suitability variables were estimated by processing standard ECRB and BMTB solutions, and the same was injected six times into a chromatographic system. The variables, like resolution, United States Pharmacopoeia (USP) plate count, and peak tailing, were estimated.^[18] The findings were represented in Table 1 and Fig. 2.

Specificity

Method specificity was determined by infusing the blank, placebo, standard, and sample solutions into a chromatographic system, and the resulting chromatograms were evaluated for interference with the excipients, degradants, and other components that may be expected to be present. Blank, standard, formulation, and placebo chromatograms were represented in Fig. 3.

Precision

The precision of the present work was assessed in terms of method and intermediate precision. The method precision (repeatability) was estimated by infusing six standard solutions and six sample solutions. Intermediate precision was evaluated by infusing six standard solutions and six sample solutions on different days by different employees on different chromatographic systems. [14,16]

The peak responses of all the chromatograms were taken, and standard deviation, % RSD, and percentage assay of sample solutions were calculated. The findings were shown in Tables 2 and 3.

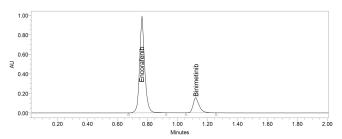


Fig. 2: System suitability chromatogram of ECRB and BMTB

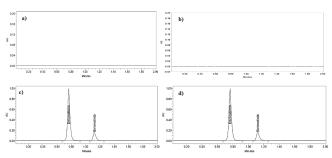


Fig. 3: Chromatograms of a) Blank; b) Placebo; c) Standard; d) Sample

Table 1: System suitability parameters for ECRB and BMTB

	ECRB			ВМТВ	ВМТВ			
S. No.	RT (min)	USP plate count	Tailing	RT (min)	USP plate count	Tailing	USP resolution	
1	0.766	2,732	1.22	1.128	3,728	1.16	5.2	
2	0.766	2,641	1.27	1.128	3,585	1.19	5	
3	0.766	2,624	1.26	1.129	3,583	1.19	5	
4	0.767	2,625	1.27	1.13	3,625	1.18	5	
5	0.767	2,686	1.24	1.13	3,548	1.18	4.9	
6	0.767	2,599	1.27	1.13	3,453	1.19	4.8	

Table 2: Repeatability results of ECRB and BMTB

Table 2. Repeatability results of Edito and Birth				
S. No.	Area of ECRB	Area of BMTB		
1.	1,246,477	243,254		
2.	1,252,281	243,393		
3.	1,251,131	242,062		
4.	1,248,204	245,139		
5.	1,242,873	242,591		
6.	1,252,694	245,621		
Mean	1,248,943	243,677		
SD	3,833.3	1,411.7		
% RSD	0.3	0.6		

SD: Standard deviation; RSD: Relative standard deviation

Table 3: Intermediate precision results of ECRB and BMTB

S. No.	Area of ECRB	Area of BMTB
1.	1,246,469	243,850
2.	1,244,094	242,664
3.	1,243,469	245,656
4.	1,237,521	242,089
5.	1,233,073	245,904
6.	1,236,125	243,739
Mean	1,240,125	243,984
SD	5,285.7	1,542.3
% RSD	0.4	0.6

SD: Standard deviation; RSD: Relative standard deviation



Table 4: Accuracy results of ECRB and BMTB

% level	Amount spiked (μg/mL)	Amount recovered (μg/mL)	% recovery	Mean % recovery	Amount spiked (μg/mL)	Amount recovered (μg/mL)	% recovery	Mean % recovery
	90	89.72457	99.69		9	8.990776	99.9	
50	90	89.71574	99.68		9	8.988794	99.88	
	90	89.68253	99.65		9	8.993215	99.92	
	180	179.4969	99.72		18	17.8281	99.04	
100	180	179.2611	99.59	99.59	18	17.91439	99.52	99.7
	100	99.08887	99.09		18	17.93818	99.66	
	270	269.1872	99.7		27	26.95213	99.82	
150	270	268.6239	99.49		27	26.97965	99.92	
	270	269.2932	99.74		27	26.88817	99.59	

Table 5: Linearity results for ECRB and BMTB

ECRB		BMTB	
Conc. (μg/mL)	Peak area	Conc. (µg/mL)	Peak area
0	0	0	0
45	322,448	4.5	60,978
90	633,298	9	123,832
135	934,969	13.5	179,592
180	1,211,063	18	235,754
225	1,536,452	22.5	300,887
270	1,805,748	27	353,700

Table 6: LOD and LOQ results for ECRB and BMTB

Analyte	LOD (μg/mL)	LOQ (μg/mL)
ECRB	0.51	1.55
BMTB	1.47	4.44

Accuracy

Method accuracy was estimated at three variable concentrations of 50, 100, and 150% levels by spiking the known amount of the drug analytes.^[17] The % recovery at each level was calculated, and the findings were represented in Table 4.

Linearity

The linearity of the developed method was evaluated by processing six different concentration levels of both ECRB and BMTB over the concentration of 45 to 270 μ g/mL and 4.5 to 27 μ g/mL. Each concentration level was processed in triplicates. ^[11,16] The linearity plots were acquired by plotting peak response (on X-axis) *vs.* concentration (on Y-axis). The results of the linearity were represented in Table 5, and Figs 4 and 5.

Limit of Detection (LOD) and Limit of Quantification (LOQ)

The LOD is the lowest quantity of drugs in a sample that can be identified but cannot be quantified exactly. LOQ is the lowest quantity of a drug in an analyte, which can be quantitatively estimated with suitable accuracy and

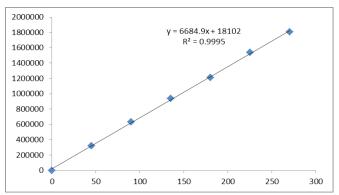


Fig. 4: Calibration curve of ECRB

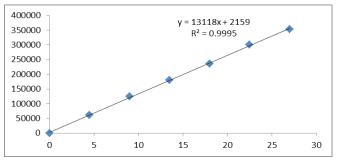


Fig. 5: Calibration curve of BMTB

precision. The LOD and LOQ values were calculated from the linearity data by utilizing the standard deviation and slope of the curve. [11,14] The resulting LOD and LOQ findings were represented in Table 6.

Robustness

The method robustness was processed by introducing small variation in the optimized liquid chromatography (LC) conditions, such as, organic phase in the mobile phase ($\pm 5\%$), flow rate (-0.27 and +0.33 mL/min), and column temperature ($\pm 5^{\circ}$ C). The findings were shown in Table 7.

Degradation Studies

Alkali Degradation Studies

To 1 mL of each ECRB and BMTB stock solution, 1 mL of 2N NaOH was added to a 10 mL volumetric flask and

kept at 60°C for 30 minutes. Further, the resulting solution was made up to the mark to get 180 and $18\,\mu g/mL$ concentrations of ECRB and BMTB, respectively. From that, 0.3 μL of the solution was infused into a UPLC system, and the resultant chromatograms were analyzed for the stability of analytes. [12] The findings were represented in Table 8 and Fig. 6.

Photolytic Stability Study

For the photolytic stability study, ECRB 1,800 µg/mL and BMTB 180 µg/mL solutions were exposed to UV-light by placing the solutions in UV cabinet for 1-day or 200 Watthours/m² in a photostability chamber. The resulting solutions were combined in a 10 mL volumetric flask and made up to the mark with diluent to get 180 and 18 µg/mL concentrations of ECRB and BMTB, respectively. From that, 0.3 µL of the solution was infused into a UPLC system, and the resultant chromatograms were analyzed for analytes' stability. The findings were represented in Table 8 and Fig. 6.

Acid Degradation Studies

To 1 mL of each ECRB and BMTB stock solution, 0.01 mL of 2N hydrochloric acid was added to a 10 mL volumetric flask and refluxed at 60°C for 30 minutes. Further, the resulting solution was made up to the mark to get 180 and 18 $\mu g/mL$ concentrations of ECRB and BMTB, respectively. From that, 0.3 μL of the solution was infused into a UPLC system, and the resultant chromatograms were analyzed for the stability of analytes. The findings were represented in Table 8 and Fig. 6.

Neutral Degradation Studies

To 1 mL of each ECRB and BMTB stock solution, 5 mL of water was added into a 10 mL volumetric flask and kept

Table 7: Robustness data for ECRB and BMTB

S. No.	Variation in LC conditions	ECRB % RSD	BMTB % RSD
1	Flow rate (-) 0.27 mL/min	0.8	0.7
2	Flow rate (+) 0.33 mL/min	0.5	0.5
3	Organic phase -5%	0.6	0.3
4	Organic phase +5%	0.6	0.6
5	Temperature at 25°C	1.1	0.9
6	Temperature at 35°C	1	0.8

for refluxing at 60°C for 1-hour. Further, the resulting solution was made up to the mark to get 180 and $18\,\mu g/mL$ concentrations of ECRB and BMTB, respectively. From that, 0.3 μL of the solution was infused into a UPLC system, and the resultant chromatograms were analyzed for the stability of analytes. The findings were represented in Table 8 and Fig. 6.

Oxidation

To 1 mL of each ECRB and BMTB stock solution, 1 mL of 20% hydrogen peroxide ($\rm H_2O_2$) was added into a 10 mL volumetric flask and kept at $60\,^{\circ}\text{C}$ for 30 minutes. Further, the resulting solution was made up to the mark to get 180 and 18 µg/mL concentrations of ECRB and BMTB, respectively. From that, 0.3 µL of the solution was infused into a UPLC system, and the resultant chromatograms were analyzed for the stability of analytes. The findings were represented in Table 8 and Fig. 6.

Dry Heat Degradation Studies

To a 10 mL volumetric flask, add 1 mL each ECRB and BMTB stock solution and monitor at 105°C for 1-hour in a hot air oven to perform the dry heat stability study. Further, the resulting solution was made up to the mark to get 180 and 18 $\mu g/mL$ concentrations of ECRB and BMTB,

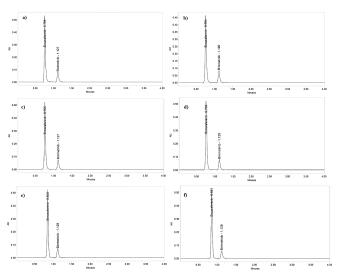


Fig. 6: Representative chromatograms of a) UV-degradation;
b) Peroxide degradation;
c) Thermal degradation;
d) Neutral degradation;
e) Acid degradation;
f) Alkali degradation

Table 8: Degradation data of ECRB and BMTB

	ECRB			ВМТВ		
	ECRB	,		DIVIID		
Type of degradation	Area	% recovered	% degraded	Area	% recovered	% degraded
Acid	1,211,851	97.47	2.53	237,330	97.34	2.66
Alkali	1,202,123	96.69	3.31	236,412	96.97	3.03
Peroxide	1,157,758	93.12	6.88	231,328	94.88	5.12
Thermal	1,218,460	98	2	236,704	97.08	2.92
UV light	1,208,181	97.18	2.82	239,749	98.33	1.67
Neutral	1,225,794	98.59	1.41	241,947	99.24	0.76



respectively. From that, $0.3\,\mu\text{L}$ of the solution was infused into a UPLC system, and the resultant chromatograms were analyzed for the stability of analytes. The findings were represented in Table 8 and Fig. 6.

DISCUSSION

After the method development trials, chromatographic parameters were optimized with mobile phase composition of $0.01 \text{N KH}_2\text{PO}_4$ buffer (pH 3.5) and acetonitrile in the ratio of 55:45~WeV/v, HSS C18 ($100 \times 2.1~\text{mm}$, 1.8~m) reverse phase column, a flow rate of 1 mL/min, column temperature of 30°C and the detection wavelength at 294 nm. The optimized method was processed for validation as per the guidelines of ICH. In the system suitability studies, the tailing factor should be < 2, the resolution must be > 2, and plate count should be > 2,000. All the suitable system parameters were passed and were within limits. Retention times of ECRB and BMTB were 0.767, 1.13, and 3.648~minutes, in that order. We did not find any additional peaks in blank and placebo at these drugs' retention times in this technique. So this technique was said to be specific.

The average area, standard deviation (SD), and % RSD were calculated for the method and intermediate precision, and the % RSD values were less than 0.6% for ECRB and BMTB. As the limit of precision was < 2 and both the precisions were passed in this analysis process. The method has a high degree of accuracy based on the mean recovery values and was 99.59 and 99.7% for ECRB and BMTB, respectively. The correlation coefficient values obtained for both the drugs were > 0.999, and it proves that the method has a high degree of linearity. The method robustness was processed by variation in the mobile phase, flow rate, column temperature, and % RSD was calculated. The resultant findings (Table 7) prove the method robustness. Further, two analytes were subjected to acid, peroxide, photolytic, alkali, neutral, and thermal degradation studies, and the results shown that the drugs were prone to degradation between 0.76 and 6.88%.

CONCLUSION

A simple, accurate, and precise method was developed for the simultaneous estimation of the ECRB and BMTB in tablet dosage form by the RP-UPLC technique. Retention times of ECRB and BMTB were found to be 0.767 and 1.13 minutes, respectively. Chromatographic elution was processed through an HSS C18 (100×2.1 mm, 1.8 m) reverse phase column, and the mobile phase composition of 0.01N KH₂PO₄ buffer (pH 3.5) and acetonitrile in the ratio of 55:45 was pumped through a column at a flow rate of 1 mL/min. Repeatability of the method was determined in the form of % RSD, and findings were 0.3 and 0.6 for ECRB and BMTB, respectively. LOD, LOQ values obtained from regression equations of ECRB and BMTB were 0.51, 1.55 mg/mL and 1.47, 4.44 mg/mL, respectively. Two

analytes were subjected to acid, peroxide, photolytic, alkali, neutral, and thermal degradation studies, and the results shown that the percentage of degradation was found between 0.76 and 6.88%. Retention times and the total run time of two drugs were decreased, and the developed method was simple and economical. So, the developed method can be adopted in industries as a regular quality control test to quantify ECRB and BMTB.

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HOW TO CITE THIS ARTICLE: Raveendranath TV, Saravanakumar RT, Male A. Stability-indicating reversed phase-ultra performance liquid chromatography method development and validation for simultaneous determination of encorafenib and binimetinib in formulation. Int. J. Pharm. Sci. Drug Res. 2020;12(5):488-494. DOI: 10.25004/IJPSDR.2020.120509

